MCDB 4312/5312 Quantitative Optical Imaging

Time:

Fall 2017 MWF, 11:30-12:20

Location:

JSCBB B331

Office hours:

Monday 1-2 PM Thursday 10-11 AM JSCBB 3rd floor, A-wing common area

Instructor:

Joel Kralj, PhD MCD Biology/BioFrontiers Institute Office: JSCBB A321 Phone: 303 492-2642 Email: <u>Quant.Optics@colorado.edu</u>

Class description:

This class is designed to explore the fundamentals of optical imaging in biology. Covered topics include an introduction to optics and microscopes, fluorescence microscopy, probes and biosensors, and image analysis. MATLAB will be taught throughout the course and used for image processing. At the end of the class, students will be able to recognize the strengths and limitations of imaging techniques as well as analyze digital images of cells and molecules.

Optional Textbook and Resources:

"Fundamentals of Light Microscopy and Electronic Imaging, 2nd Edition" Douglas B. Murphy, Michael W. Davidson ISBN: 978-0-471-69214-0 This is a useful text with information on the nature of light, specific imaging modalities, and basics on image processing.

"From Photon to Neuron: Light, Imaging, Vision" Philip Nelson ISBN: 978-1-400-885-480 This is a very interesting, but much more mathematically inclined, book that covers the nature of light and how biological systems deal with light as a physical quantity.

Web resources:

General Microscopy

Microscopy U - <u>http://www.microscopyu.com/</u> (Nikon) Microscopy Resource Center - <u>http://www.olympusmicro.com/</u> (Olympus) Zeiss Campus - <u>http://zeiss-campus.magnet.fsu.edu/index.html</u> (Zeiss)

Matlab

Matlab Onramp: https://matlabacademy.mathworks.com/

Intro to matlab (PDF) https://www.google.com/url?sa=t&rct=j&q=&esrc=s&source=web&cd=3&ved=0ahUKEwjYm8 jdlbfOAhUX9mMKHVwaBywQFgg1MAI&url=https%3A%2F%2Fwww.mccormick.northwest ern.edu%2Fdocuments%2Fstudents%2Fundergraduate%2Fintroduction-tomatlab.pdf&usg=AFQjCNEuH8v-uDmcqfEDIAjpsUV3yV5ww&sig2=FfbgSmx_AxHaZyOrnBY2FQ&cad=rja

Basic image processing - <u>http://www.mathworks.com/help/images/getting-started-with-image-processing-toolbox.html</u>

Image processing - http://www.getreuer.info/tutorials/matlabimaging

Graduate Credit:

Graduate students can also be enrolled in a 1 credit practical class. This class meets weekly for 1 hour to use microscopes in the BioFrontiers Advanced Imaging Core to answer fundamental questions about optics and microscopy and is led by Joe Dragavon. Students will have additional homeworks in the section, and will be required to design, perform, and analyze measurements on live cells.

Grading:

10% Class attendance/discussion 40% Homeworks 15% Midterm 1 – 2017-09-27 15% Midterm 2 – 2017-11-01 20% Final exam – 2017-12-19 (4:30 – 7:00)

Topics:

Unit 1: Light and optics

- 1. Fundamentals of Light and color
- 2. Intro to refraction, lenses and ray optics
- 3. Images, virtual images, and magnification

- 4. Reflection, diffraction and microscope components
- 5. Diffraction, spatial resolution, and NA
- 6. Aberrations and corrections
- 7. Light sources, speckle

Unit 2: Microscopy contrast mechanisms

- 8. Phase contrast and dark field microscopy
- 9. Polarized light and microscopy
- 10. DIC microscopy
- 11. Fundamentals of fluorescence
- 12. Dyes, proteins, and filters
- 13. Limitations of fluorescence and practical applications

Unit 3: Techniques in fluorescence

- 14. Imaging in cells: FRET and TIRF
- 15. Imaging in cells: FRAP, photoactivation, and FLIM
- 16. Principles of biosensors
- 17. Sensing pH, calcium, voltage, ATP etc...
- 18. High content screening
- 19. Confocal optics
- 20. Spinning disk confocal
- 21. Superresolution techniques 1
- 22. Superresolution techniques 2
- 23. Multiphoton fluorescence
- 24. Adaptive optics
- 25. Imaging live cells, light sheet, Bessel
- 26. Imaging live cells, light field, ultra wide field

Unit 4: Image detection and analysis

- 27. Fundamentals of digital detectors
- 28. Noise and sensitivity
- 29. Image analysis: Contrast, gamma correction, and histograms
- 30. Image analysis: Filters and segmentation 1
- 31. Image analysis: Filters and segmentation 2
- 32. Image analysis: Time course analysis
- 33. Image analysis: Cell tracking
- 34. Image analysis: Cell tracking 2

Administrative:

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http://www.colorado.edu/policies/classbehavior.html and at http://www.colorado.edu/studentaffairs/judicialaffairs/code.html#student_code

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